# Selective exploitation of early running fish may induce genetic and phenotypic changes in Atlantic salmon 

S. Consuegra*, C. García de Leániz $\dagger \dagger$, A. Serdio§ and E. Verspoor ${ }^{\boldsymbol{T}}$<br>*University of St Andrews, Gatty Marine Laboratory, St Andrews, Fife KY16 8LB, Scotland, U.K., †University of Wales Swansea, School of Biological Sciences, Swansea SA2 8PP, U.K., §Centro Ictiológico de Arredondo, Consejería de Ganadería, Agricultura y Pesca, 39813 Arredondo, Cantabria, Spain and【Fisheries Research Service, Freshwater Laboratory, Pitlochry, Perthshire PH16 5LJ, Scotland, U.K.


#### Abstract

Genetic evidence for the selective exploitation by anglers of early running fish was examined in four Iberian Atlantic salmon populations using protein and mtDNA markers. The populations studied had been exploited exclusively by anglers since 1949 during a fixed fishing season that ran approximately from March to July. Genetic variation at six protein loci was small and was accounted for by the $M E P-2^{*}$ and $M D H-3,4^{*}$ polymorphisms, which generally remained stable over time and were in Castle-Hardy-Weinberg equilibrium during the fishing season. Early running fish that had spent multiple winters at sea (MSW) generally had higher frequencies of the common $M E P-2^{*}$ (100) allele than did late running, one sea winter (1SW) grilse that were significantly smaller and tended to escape the fishery. Spawners differed from angler caught fish in their mtDNA frequencies and consistently had a lower seaage and a smaller body size. Spawners also smolted at an older age and displayed lower frequencies of the $M E P-2^{*}(100)$ allele in three of the four populations studied. These results suggest that in these rivers anglers selectively exploit a distinct component of the population and inadvertently cause a differential mortality of genetic types that is likely to be detrimental to population viability. © 2005 The Fisheries Society of the British Isles


Key words: Atlantic salmon; conservation; isozymes; mtDNA; population structuring; selective exploitation.

## INTRODUCTION

Many commercially valuable fish species, including salmonids, are exploited seasonally to give protection from fishing at vulnerable life history stages or during spawning, or a better chance to reproduce. The side effects of seasonal fishing closures, however, have seldom been considered. For example, despite clear indications that salmonids are structured both spatially and temporally (Hansen \& Jonsson, 1991; Adams et al., 1994; Ferguson et al., 1995), some

[^0]populations of Atlantic salmon Salmo salar L. continue to be exploited in many rivers during only a few months of the year, typically from early spring until summer. Consequently, some individuals or components of the population may be subjected to higher mortality rates than others, and a certain degree of artificial selection may occur.

In the case of Atlantic salmon, adults that enter the rivers in the spring are often dominated by large multi sea-winter (MSW) females, whereas fish that enter rivers in late summer and fall are often dominated by male-biased, one seawinter grilse (Gardner, 1976; Shearer, 1992). Since traits such as age at maturity or timing of entry into the rivers are partially inherited and likely to be adaptive (Hansen \& Jonsson, 1991; Taylor, 1991; Stewart et al., 2002), there is considerable scope for differential mortality of genetic types and, hence, a definite risk of inadvertent artificial selection by anglers.

Exploited salmonid populations have been shown to respond to differential mortality in several ways. For example, when the fishery is concentrated upon the largest individuals (Gee \& Milner, 1980), fish populations may respond by decreasing their average body size or by maturing at an earlier sea-age (Thorpe, 1993). Other phenotypic responses attributed to exploitation in Atlantic salmon include a shift towards delayed entry into rivers (García de Leániz et al., 1992) or a greater incidence of sexually mature male parr (Caswell et al., 1984). In contrast, it has proved much more difficult to detect explicit genetic effects that are attributable to fisheries perhaps due to the low resolution of the molecular markers employed, which may only cover a small part of the genome (Conover \& Munch, 2002). Consequently, the role of fishing in changing the genetic structure of fish populations remains controversial (Smith et al., 1991; Policansky, 1993; Hutchings, 2001; Law, 2001).

In the present study, protein and mitochondrial DNA variation were examined in relation to population structuring and differential exploitation in endangered Atlantic salmon populations from Cantabria (N. Spain). Evidence for selective exploitation was assessed by comparing genetic and phenotypic traits between (1) early and late running fish and (2) fish caught by anglers and fish that survived to spawn.

## MATERIALS AND METHODS

## STUDY RIVERS AND COLLECTION OF SAMPLES

The four small populations studied (rivers Asón, Pas, Nansa, and Deva) have been exploited exclusively by rod and line anglers since 1949 over a fairly constant fishing season, typically extending from mid-March to mid-July (García de Léaniz \& Martinez, 1988; García de Leániz et al., 1989). Average yearly rod and line catches during 19882000 ranged from $48 \pm 61 \cdot 5$ for the River Deva to $108 \pm 135 \cdot 9$ for the River Pas. Samples of muscle tissue and adipose fins were obtained from nearly all fish caught by anglers during the period 1988-2000, as well as from a sample of spawners and adults that escaped the fishery during the same period. Liver tissue was collected from a smaller sub-sample of fish. Most spawners were collected in adult fish traps located in the same, short ( $<40 \mathrm{~km}$ ) accessible reaches of the main rivers where angling took place, or by electrofishing. Data for the River Nansa indicated that there was no size difference between spawners caught by electrofishing and spawners caught in fish traps (MannWhitney, $P=0.944$ ). Fork length, $L_{\mathrm{F}}(\mathrm{cm})$ and wet mass $(\mathrm{g})$ were recorded from most
fish and samples of scales taken for age determination. Fish were classified as grilse (one winter at sea) or multiple sea winter (MSW; 2-3 winters at sea) based on scale reading or (when scales were not available or could not be read) according to their $L_{\mathrm{F}}$, using 70 cm as the cut-off point between both age groups. Hatchery-reared fish (all identified by finclipping since 1992) were excluded from analyses, as these are known to differ from wild individuals (Verspoor, 1988, 1994).

## PROTEIN ELECTROPHORESIS

The following isozymes were screened in skeletal muscle: NADP+ dependent malic enzyme (MEP-2; EC.1.1.1.40), malate dehydrogenase (MDH-3,4, EC.1.1.1.37), phosphoglucomutase (PGM-1, EC.5.4.2.2) and glucose-6-phosphate isomerase (GPI-1,2, EC.5.3.1.9). Lactate dehydrogenase (LDH-4, EC.1.1.1.27) and isocitrate dehydrogenase (IDHP-3, EC.1.1.1.42) were screened in liver. These loci were examined because previously they had been shown to be polymorphic in Iberian salmon populations (García de Leániz et al., 1989; Sánchez et al., 1991, 1996; Morán et al., 1994). Horizontal starch gel electrophoresis was carried out according to Verspoor (1988) using the buffer system citrate-aminomorpholine described in Clayton \& Tretiak (1972), adjusted to pH 6.6 (Verspoor \& Cole, 1989) and the Tris-citrate-borate system as described in Verspoor (1988). Protein designation follows Shaklee et al. (1989). The most common allele is designated 100 and other alleles were scored according to their relative electrophoretic mobility. Loci that proved to be monomorphic in a preliminary screening were excluded from the analysis. Departure from Castle-Hardy-Weinberg equilibrium was tested using the GENEPOP package (v. 3.2, Raymond \& Rousset, 1995). Homogeneity of allele frequencies among year classes was tested using the Fisher's exact test. Differences in genotype frequencies between grilse and MSW fish, as well as between spawners and angler caught fish, were tested for statistical significance with an unbiased log-likelihood test.

## mtDNA ANALYSIS

Total DNA was extracted from the adipose fins (Taggart et al., 1992) of 673 wild adult salmon collected during 1996-2000. One $\mu$ l of the extracted DNA was used to amplify a 1400 bp fragment in the ND1 region with the 16sRNA (Palumbi, 1996) and ND1 (Cronin et al., 1993) primers, followed by digestion with the restriction enzymes HaeIII, HinfI, RsaI and DraI (Verspoor et al., 1999). Differences in haplotype composition between spawners and fish caught by anglers were examined using the MONTE program in the REAP software package (McElroy et al., 1991).

## RESULTS

## PROTEIN VARIATION

Of the six protein loci that were screened, only two loci (MEP-2* and MDH$3,4^{*}$ ) were found to be variable in Cantabrian populations (Table I). All rivers had high frequencies of the common $* 100$ allele for both loci, although no significant association was detected between MEP-2* and MDH-3,4* genotypes ( $G=2 \cdot 754$, d.f. $=2, n=863, P=0 \cdot 252$ ). Heterogeneity analysis between the four populations, on the other hand, indicated the existence of spatial structure at the two polymorphic loci ( $G$-test on allele counts; $M E P-2^{*}, \chi^{2}=21 \cdot 80$, d.f. $=3, P<0.001 ; M D H-3,4^{*}, \chi^{2}=56 \cdot 48$, d.f. $=3, P<0 \cdot 001$ ).

Table I. Protein variation among wild adult Atlantic salmon returning to Cantabrian rivers during the period 1988-2000, excluding known hatchery reared fish (identified by fin-clipping). Sample size (in parentheses) and frequency of the common allele are shown for each locus

| River | Locus |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | MEP-2* | GPI-1,2* | LDH-4* | IDHP-3* | MDH-3,4* | PGM-1* |
| Asón | 0.917* | $1 \cdot 000$ | $1 \cdot 000$ | $1 \cdot 000$ | $0 \cdot 989$ | $1 \cdot 000$ |
|  | (682) | (24) | (24) | (24) | (417) | (71) |
| Pas | $0 \cdot 966$ | $1 \cdot 000$ | $1 \cdot 000$ | $1 \cdot 000$ | $0 \cdot 909$ | $1 \cdot 000$ |
|  | (294) | (4) | (4) | (4) | (121) | (122) |
| Nansa | 0.939* | $1 \cdot 000$ | $1 \cdot 000$ | $1 \cdot 000$ | $0 \cdot 949$ | $1 \cdot 000$ |
|  | (469) | (13) | (13) | (13) | (236) | (106) |
| Deva | 0.954* | 1.000 | $1 \cdot 000$ | $1 \cdot 000$ | $0 \cdot 891$ | $1 \cdot 000$ |
|  | (346) | (7) | (7) | (7) | (110) | (86) |

*Significant deviations from CHW.

## DEPARTURES FROM CASTLE-HARDY-WEINBERG EQUILIBRIUM

All year classes that contributed to the rod and line fisheries were found to be in Castle-Hardy-Weinberg equilibrium for both loci in the four rivers studied (Tables II and III). Heterozygote deficiencies were detected for $M E P-2^{*}$ in the River Asón in the 1993 year class $(P=0 \cdot 026)$ and for $M D H-3,4^{*}$ in the River Nansa in the 1996 year class $(P=0 \cdot 010)$, but these were not significant after adjusting for multiple testing using a Bonferroni correction.

## TEMPORAL STABILITY IN GENOTYPE FREQUENCIES

Genotype frequencies during the fishing season were stable over time for both $M E P-2^{*}$ (Table II) and $M D H-3,4^{*}$ (Table III). No significant heterogeneity in genotype frequencies was found between year classes in the rivers Asón, Nansa and Deva. Only the River Pas had significant temporal heterogeneity for $M E P$ 2* genotype frequencies, due in part to lack of variation in two consecutive year classes (1995 and 1996, Table II).

## INFLUENCE OF MEP-2* ON SEA-AGE AND FISH SIZE

Analysis of sea-age (Table IV) indicated that the MEP-2* genotype was associated with the time adults spent at sea in the River Asón $(P=0.003)$, River Nansa ( $P=0 \cdot 040$ ) and perhaps also in the River Deva $(P=0 \cdot 052)$. In every case, analysis of the standardized residuals indicated that fish that were homozygous for the *100 allele tended to mature later (i.e. they were MSW salmon) than did fish with the *125 allele that were more likely to become grilse. Combined probabilities across rivers using Fisher's exact test indicated that the observed association between $M E P-2^{*}$ genotype and sea-age is unlikely to be an artifact ( $\chi^{2}=24 \cdot 62$, d.f. $=8, P=0 \cdot 0018$ ).

Table II. Distribution of $M E P-2^{*}$ genotypes among wild adults sampled in the rod and line fisheries (1988-2000) by year classes. Temporal stability in genotype frequencies and departures from Castle-Hardy-Weinberg (CHW) equilibrium are indicated

| Year class | MEP-2* genotype |  |  | Total | $F_{\text {IS }}$ | CHW | Frequency (100) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 100/100 | 100/125 | 125/125 |  |  |  |  |
| River Asón, $P=0.227$ |  |  |  |  |  |  |  |
| 1983 | 1 | 0 | 0 | 1 | - | - | - |
| 1984 | 18 | 4 | 0 | 22 | -0.080 | NS | 0.925 |
| 1985 | 183 | 19 | 4 | 206 | $+0.249$ | NS | 0.934 |
| 1986 | 22 | 1 | 0 | 23 | -0.012 | NS | 0.979 |
| 1987 | 16 | 4 | 0 | 20 | -0.111 | NS | $0 \cdot 900$ |
| 1988 | 15 | 4 | 1 | 20 | $+0 \cdot 216$ | NS | $0 \cdot 850$ |
| 1989 | 1 | 0 | 0 | 1 | - | - | - |
| 1991 | 4 | 1 | 0 | 5 | $0 \cdot 000$ | NS | $0 \cdot 900$ |
| 1992 | 27 | 6 | 1 | 34 | $+0 \cdot 165$ | NS | $0 \cdot 882$ |
| 1993 | 73 | 18 | 5 | 96 | $+0.252$ | NS | $0 \cdot 854$ |
| 1994 | 27 | 4 | 0 | 31 | -0.053 | NS | 0.935 |
| 1995 | 21 | 1 | 1 | 23 | $+0.656$ | NS | $0 \cdot 935$ |
| 1996 | 19 | 3 | 0 | 22 | $-0 \cdot 050$ | NS | 0.932 |
| 1997 | 3 | 0 | 0 | 3 | - | - | - |
| River Nansa, $P=0.835$ |  |  |  |  |  |  |  |
| 1984 | 10 | 3 | 0 | 13 | -0.130 | NS | $0 \cdot 885$ |
| 1985 | 34 | 1 | 0 | 35 | -0.010 | NS | $0 \cdot 986$ |
| 1986 | 19 | 2 | 1 | 22 | $+0.450$ | NS | $0 \cdot 909$ |
| 1987 | 2 | 0 | 0 | 2 | $0 \cdot 000$ | - | - |
| 1992 | 5 | 1 | 0 | 6 | -0.091 | NS | $0 \cdot 917$ |
| 1993 | 57 | 4 | 1 | 62 | $+0.299$ | NS | $0 \cdot 952$ |
| 1994 | 29 | 3 | 0 | 32 | -0.049 | NS | 0.953 |
| 1995 | 21 | 3 | 0 | 24 | -0.067 | NS | 0.938 |
| 1996 | 18 | 2 | 0 | 20 | $-0.053$ | NS | 0.950 |
| 1997 | 27 | 3 | 1 | 31 | $+0.347$ | NS | 0.919 |
| 1998 | 10 | 1 | 0 | 12 | -0.091 | NS | $0 \cdot 917$ |
| River Pas, $P=0.010$ |  |  |  |  |  |  |  |
| 1991 | 1 | 0 | 0 | 1 | - | - | - |
| 1992 | 19 | 1 | 0 | 20 | -0.026 | NS | 0.975 |
| 1993 | 70 | 7 | 0 | 77 | -0.048 | NS | 0.955 |
| 1994 | 11 | 3 | 1 | 15 | $+0 \cdot 280$ | NS | 0.833 |
| 1995 | 16 | 0 | 0 | 16 | $0 \cdot 000$ | NS | 1.000 |
| 1996 | 16 | 0 | 0 | 16 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1997 | 49 | 1 | 0 | 50 | -0.010 | NS | $0 \cdot 990$ |
| River Deva, $P=0.205$ |  |  |  |  |  |  |  |
| 1992 | 6 | 2 | 0 | 8 | $-0 \cdot 143$ | NS | 0.875 |
| 1993 | 181 | 14 | 2 | 197 | $+0 \cdot 187$ | NS | 0.954 |
| 1994 | 26 | 0 | 0 | 26 | $0 \cdot 000$ | NS | 1.000 |
| 1995 | 4 | 0 | 0 | 4 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1996 | 7 | 1 | 0 | 8 | -0.067 | NS | $0 \cdot 937$ |
| 1997 | 65 | 4 | 0 | 69 | -0.030 | NS | 0.971 |

Table III. Distribution of $M D H-3,4^{*}$ genotypes among wild adults sampled in the rod and line fisheries by year classes. Temporal stability in genotype frequencies and departures from Castle-Hardy-Weinberg (CHW) equilibrium are indicated

| Year class | MDH-3,4* genotype |  |  | Total | $F_{\text {IS }}$ | CHW | Frequency (100) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 100/100 | 100/80 | 80/80 |  |  |  |  |
| River Asón, $P=0 \cdot 164$ |  |  |  |  |  |  |  |
| 1983 | 1 | 0 | 0 | 1 | - | - | - |
| 1984 | 22 | 0 | 0 | 22 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1985 | 205 | 2 | 0 | 207 | -0.002 | NS | 0.995 |
| 1986 | 23 | 0 | 0 | 23 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1987 | 14 | 0 | 0 | 14 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1988 | 10 | 0 | 0 | 10 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1993 | 9 | 0 | 0 | 9 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1994 | 25 | 0 | 0 | 25 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1995 | 20 | 2 | 0 | 22 | -0.024 | NS | 0.955 |
| 1996 | 21 | 1 | 0 | 22 | $-0 \cdot 000$ | NS | 0.977 |
| 1997 | 3 | 0 | 0 | 3 | - | - | - |
| River Nansa, $P=0.053$ |  |  |  |  |  |  |  |
| 1984 | 13 | 0 | 0 | 13 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1985 | 35 | 0 | 0 | 35 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1986 | 22 | 0 | 0 | 22 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1987 | 2 | 0 | 0 | 2 | - | - | - |
| 1994 | 2 | 0 | 0 | 2 | - | - | - |
| 1995 | 7 | 1 | 0 | 8 | -0.067 | NS | 0.938 |
| 1996 | 17 | 1 | 2 | 20 | $+0.771$ | NS | $0 \cdot 875$ |
| 1997 | 25 | 6 | 0 | 31 | -0.107 | NS | 0.903 |
| 1998 | 9 | 3 | 0 | 12 | $-0.143$ | NS | $0 \cdot 875$ |
| River Pas, $P=0 \cdot 180$ |  |  |  |  |  |  |  |
| 1996 | 4 | 4 | 0 | 8 | -0.333 | NS | 0.750 |
| 1997 | 45 | 10 | 0 | 55 | -0.102 | NS | 0.909 |
| River Deva, $P=0.602$ |  |  |  |  |  |  |  |
| 1992 | 0 | 1 | 0 | 1 | - | - | - |
| 1993 | 10 | 3 | 0 | 13 | -0.130 | NS | $0 \cdot 885$ |
| 1994 | 4 | 1 | 0 | 5 | -0.111 | NS | $0 \cdot 900$ |
| 1995 | 4 | 0 | 0 | 4 | $0 \cdot 000$ | NS | $1 \cdot 000$ |
| 1996 | 5 | 2 | 0 | 7 | -0.167 | NS | $0 \cdot 857$ |
| 1997 | 49 | 16 | 0 | 65 | $-0.133$ | NS | $0 \cdot 877$ |

To dissociate the effect of the $M E P-2^{*}$ genotype on the sea-age and size of returning adults, and due also to the small number of $M E P-2^{*} 125 / 125$ homozygotes present in Cantabrian populations, data from the four rivers were pooled together. Fish that were homozygous for the MEP-2* 100 allele were significantly larger (Kruskal-Wallis, $P=0.001$ ), heavier (Kruskal-Wallis, $P=0 \cdot 002$ ) and remained longer at sea (Kruskal-Wallis, $P<0 \cdot 001$ ) than fish that were homozygous for the $M E P-2^{*} 125$ allele, whereas heterozygotes were intermediate in size and sea-age (Figs 1 and 2). Within each age class, however, no difference existed between $M E P-2^{*}$ genotype and $L_{\mathrm{F}}$ for either grilse

Table IV. Distribution of $M E P-2^{*}$ and $M D H-3,4^{*}$ genotypes among wild grilse and MSW adults sampled during the 1988-2000 runs. Probabilities refer to a Fisher's exact test for differences in allelic frequencies

| River/Age Class | MEP-2* genotype |  |  | Total | Frequency (100) |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | 100/100 | 100/125 | 125/125 |  |  |
| River Asón, $P=0.003$ |  |  |  |  |  |
| Grilse | 75 | 17 | 7 | 99 | $0 \cdot 843$ |
| MSW | 477 | 64 | 8 | 549 | 0.927 |
| River Pas, $P=0.722$ |  |  |  |  |  |
| Grilse | 30 | 1 | 0 | 31 | 0.984 |
| MSW | 242 | 17 | 1 | 260 | $0 \cdot 963$ |
| River Nansa, $P=0.040$ |  |  |  |  |  |
| Grilse | 147 | 21 | 4 | 172 | 0.916 |
| MSW | 264 | 18 | 3 | 285 | $0 \cdot 958$ |
| River Deva, $P=0.052$ |  |  |  |  |  |
| Grilse | 28 | 1 | 2 | 31 | $0 \cdot 919$ |
| MSW | 291 | 21 | 3 | 315 | 0.957 |
| MDH-3,4* genotype |  |  |  |  |  |
| River/Age Class | 100/100 | 100/80 | 80/80 | Total | Frequency (100) |
| River Asón, $P=0.607$ |  |  |  |  |  |
| Grilse | 58 | 2 | 0 | 60 | $0 \cdot 967$ |
| MSW | 349 | 7 | 0 | 356 | $0 \cdot 980$ |
| River Pas, $P=0 \cdot 181$ |  |  |  |  |  |
| Grilse | 13 | 3 | 1 | 17 | $0 \cdot 853$ |
| MSW | 85 | 17 | 0 | 102 | $0 \cdot 917$ |
| River Nansa, $P=0.423$ |  |  |  |  |  |
| Grilse | 70 | 9 | 0 | 79 | 0.943 |
| MSW | 134 | 11 | 2 | 147 | 0.949 |
| River Deva, $P=0 \cdot 121$ |  |  |  |  |  |
| Grilse | 11 | 0 | 0 | 11 | $1 \cdot 000$ |
| MSW | 75 | 24 | 0 | 99 | 0.879 |

(Kruskal-Wallis, $P=0.589$ ) or MSW (Kruskal-Wallis, $P=0 \cdot 609$ ). This suggested that the $M E P-2^{*}$ genotype was primarily associated with age at maturity and that this, in turn, determined the size attained by returning fish. Thus, average grilsing rates in the four populations increased from $17 \cdot 3 \%$ for $M E P$ 2* 100/100 to $47.9 \%$ for $M E P-2^{*} 125 / 125$, while the incidence of grilse among $M E P-2^{*} 100 / 125$ was intermediate ( $21 \cdot 2 \%$ ).

No significant association was found between the $M E P-2^{*}$ genotype and freshwater age (Kruskal-Wallis, $P=0 \cdot 208$ ) or between the $M D H-3,4^{*}$ genotype (pooling together fish with the $* 80$ allele) and $L_{\mathrm{F}}$ (Kruskal-Wallis, $P=0.367$ ), sea-age (Kruskal-Wallis, $P=0 \cdot 856$ ), or freshwater age (Kruskal-Wallis, $P=0.683$ ) of returning adults.


Fig. 1. Relationship between $M E P-2^{*}$ genotype and variation in (a) $L_{\mathrm{F}}(\mathrm{cm})$ and (b) wet mass (kg) of adult salmon returning to Cantabrian streams (including both fish caught by anglers and spawners), tested for statistical significance by the Kruskal-Wallis $H$-statistic. Box and whisker plots show median values, first and third quartiles ( $50 \%$ of values) and extreme values ( O ). Notches extend to $95 \%$ CIs around the median, shown in numbers. Sample size in parentheses.


Fig. 2. Relationship between $M E P-2^{*}$ genotype and variation in (a) sea age (yr) and (b) freshwater age (yr) of adult salmon (fishery + spawners) returning to Cantabrian streams (means, s.d.), tested for statistical significance by the Kruskal-Wallis $H$-statistic. Sample size in parentheses.

## PHENOTYPIC AND GENETIC DIFFERENCES BETWEEN SPAWNERS AND FISH CAUGHT BY ANGLERS

Data for Cantabrian populations indicated that the average size and sea-age of returning adults decreased markedly as the season progressed (Fig. 3). Consequently, salmon caught by anglers during the fishing season (mid-March to mid-July) were significantly larger and matured at an older sea-age than salmon that survived to spawn in all four populations (Table V). Compared to


FIG. 3. Monthly variation (mean, s.D.) in $L_{\mathrm{F}}(\mathrm{cm}, n=1720)$ and sea-age $(\mathrm{yr}, \mathrm{n}=1600)$ of wild adult salmon (fishery + spawners) returning to Cantabrian rivers during the period 1988-2000. The extent of the angling season is indicated.
angler caught fish, spawners also tended to smolt at a significantly older age in three of the four populations.

Genetic differences between angler caught fish and spawners were also evident at isozyme and mtDNA levels, which again indicated the existence of selective exploitation by the fishery. Thus spawners had a higher frequency of the MEP2* 125 allele than did fish caught by anglers in three of the four populations studied. Although such differences were only statistically significant in the River
Table V. Demographic and genetic differences between spawners and fish caught by anglers in four Iberian Atlantic salmon populations (means $\pm$ s.e.; sample size in parentheses). Probabilities refer to the null hypotheses of no difference in age or body size (tested by the nonparametric Kruskal-Wallis) or in allele/haplotypic frequencies (tested by a Fisher's exact test)

|  |  |  |  | MEP-2* |  |  |  | MDH-3,4* |  |  |  | mtDNA haplotype HaeIII, HinfI, RsaI, DraI |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Freshwater age (year) | Sea-age (year) | $\begin{aligned} & L_{\mathrm{F}} \\ & (\mathrm{~cm}) \end{aligned}$ | $\begin{gathered} 1001 \\ 100 \end{gathered}$ | $\begin{array}{r} 1001 \\ 125 \end{array}$ | $\begin{gathered} 125 / \\ 125 \end{gathered}$ | Frequency <br> (100) | $\begin{gathered} 1001 \\ 100 \end{gathered}$ | $\begin{gathered} 100 / \\ 80 \end{gathered}$ | $\begin{gathered} 801 \\ 80 \end{gathered}$ | Frequency <br> (100) | AAAB | AABB | DABB | DBBB | DAAB | Frequency (AAAB) |
| River Asón |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Fishery | $\begin{gathered} 1 \cdot 25 \pm 0 \cdot 02 \\ (517) \end{gathered}$ | $\begin{gathered} 1 \cdot 93 \pm 0.01 \\ \quad(523) \end{gathered}$ | $\begin{gathered} 77 \cdot 0 \pm 0 \cdot 23 \\ (552) \end{gathered}$ | 489 | 67 | 12 | $\begin{aligned} & 0.920 \\ & (568) \end{aligned}$ | 356 | 6 | 0 | $\begin{aligned} & 0 \cdot 992 \\ & (362) \end{aligned}$ | 59 | 26 | 1 | 0 | 0 | $\begin{gathered} 0 \cdot 686 \\ (86) \end{gathered}$ |
| Spawners | $\begin{gathered} 1.36 \pm 0.06 \\ (76) \end{gathered}$ | $\begin{gathered} 1 \cdot 40 \pm 0.06 \\ (76) \end{gathered}$ | $\begin{gathered} 70 \cdot 3 \pm 0 \cdot 80 \\ (101) \end{gathered}$ | 95 | 16 | 3 | $\begin{aligned} & 0 \cdot 903 \\ & (114) \end{aligned}$ | 52 | 3 | 0 | $\begin{gathered} 0 \cdot 973 \\ (55) \end{gathered}$ | 58 | 8 | 0 | 0 | 0 | $\begin{gathered} 0 \cdot 879 \\ (66) \end{gathered}$ |
| $P$ | $0 \cdot 045$ | $0 \cdot 000$ | $0 \cdot 000$ |  |  |  | 0.749 |  |  |  | $0 \cdot 118$ |  |  |  |  |  | $0 \cdot 007$ |
| River Pas |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Fishery | $\begin{gathered} 1 \cdot 19 \pm 0 \cdot 03 \\ (204) \end{gathered}$ | $\begin{gathered} 2 \cdot 00 \pm 0 \cdot 01 \\ \quad(207) \end{gathered}$ | $\begin{gathered} 78 \cdot 7 \pm 0 \cdot 28 \\ (229) \end{gathered}$ | 206 | 14 | 1 | $\begin{aligned} & 0 \cdot 964 \\ & (221) \end{aligned}$ | 62 | 17 | 0 | $\begin{gathered} 0.892 \\ (79) \end{gathered}$ | 95 | 0 | 3 | 0 | 1 | $\begin{gathered} 0 \cdot 960 \\ (99) \end{gathered}$ |
| Spawners | $\begin{gathered} 1.37 \pm 0.07 \\ (60) \end{gathered}$ | $\begin{gathered} 1 \cdot 62 \pm 0.07 \\ (61) \end{gathered}$ | $\begin{gathered} 72 \cdot 2 \pm 1 \cdot 02 \\ (70) \end{gathered}$ | 69 | 4 | 0 | $0.973$ <br> (73) | 38 | 3 | 1 | $\begin{gathered} 0 \cdot 940 \\ (42) \end{gathered}$ | 73 | 6 | 4 | 0 | 0 | $\begin{gathered} 0 \cdot 880 \\ (83) \end{gathered}$ |
| $P$ | $0 \cdot 007$ | $0 \cdot 000$ | $0 \cdot 000$ |  |  |  | 1.000 |  |  |  | $0 \cdot 251$ |  |  |  |  |  | $0 \cdot 015$ |
| River Nansa |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Fishery | $\begin{gathered} 1 \cdot 11 \pm 0 \cdot 02 \\ (259) \end{gathered}$ | $\begin{gathered} 1 \cdot 86 \pm 0 \cdot 03 \\ (261) \end{gathered}$ | $\begin{gathered} 78 \cdot 1 \pm 0 \cdot 47 \\ (285) \end{gathered}$ | 258 | 24 | 3 | $\begin{aligned} & 0.947 \\ & (285) \end{aligned}$ | 133 | 12 | 2 | $\begin{aligned} & 0.946 \\ & (147) \end{aligned}$ | 105 | 6 | 1 | 0 | 0 | $\begin{aligned} & 0 \cdot 929 \\ & (112) \end{aligned}$ |
| Spawners $P$ | $\begin{gathered} 1 \cdot 23 \pm 0.03 \\ (159) \end{gathered}$ | $\begin{gathered} 1.30 \pm 0.04 \\ (159) \end{gathered}$ | $\begin{gathered} 67 \cdot 9 \pm 0.57 \\ (168) \end{gathered}$ | 161 | 19 | 4 | $0.927$ <br> (184) | 81 | 8 | 0 | $\begin{gathered} 0 \cdot 955 \\ (89) \end{gathered}$ | 101 | 22 | 4 | 0 | 0 | $\begin{aligned} & 0.795 \\ & (127) \end{aligned}$ |
| River Deva |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Fishery | $\begin{gathered} 1 \cdot 16 \pm 0 \cdot 02 \\ (313) \end{gathered}$ | $\begin{gathered} 1.97 \pm 0.01 \\ \quad(315) \end{gathered}$ | $\begin{gathered} 79 \cdot 9 \pm 0 \cdot 27 \\ (318) \end{gathered}$ | 297 | 21 | 2 | $\begin{aligned} & 0 \cdot 961 \\ & (320) \end{aligned}$ | 73 | 23 | 0 | $\begin{gathered} 0 \cdot 880 \\ (96) \end{gathered}$ | 78 | 0 | 1 | 0 | 0 | $\begin{gathered} 0.987 \\ (79) \end{gathered}$ |
| Spawners | $\begin{gathered} 1 \cdot 15 \pm 0 \cdot 07 \\ (26) \end{gathered}$ | $\begin{gathered} 1.31 \pm 0.09 \\ \quad(26) \end{gathered}$ | $\begin{gathered} 67 \cdot 6 \pm 1 \cdot 34 \\ (26) \end{gathered}$ | 22 | 1 | 3 | $\begin{gathered} 0 \cdot 865 \\ (26) \end{gathered}$ | 13 | 1 | 0 | $\begin{gathered} 0 \cdot 964 \\ (14) \end{gathered}$ | 16 | 3 | 1 | 1 | 0 | $\begin{gathered} 0.762 \\ (21) \end{gathered}$ |
| $P$ | $0 \cdot 971$ | $0 \cdot 000$ | $0 \cdot 000$ |  |  |  | $0 \cdot 009$ |  |  |  | $0 \cdot 329$ |  |  |  |  |  | $0 \cdot 002$ |

Deva ( $P=0.009$ ) and River Nansa ( $P=0.013$; Table V), they were significant if considered collectively across the four rivers (Fisher's exact test for combined probabilities, $\chi^{2}=18.68$, d.f. $=8, P=0.016$ ). No such difference between spawners and angler caught fish was evident with respect to $M D H-3,4^{*}$ allele frequencies (Fisher's test of combined probabilities, $\chi^{2}=10 \cdot 01$, d.f. $=8$, $P=0.251$ ).

Data from mitochondrial DNA indicated that there were significant genetic differences between angler caught fish and spawners in all four populations (Table V). In three of the populations, fish caught by anglers had a higher frequency of the most common AAAB haplotype, while in the River Asón the opposite trend (lower frequency of the common haplotype amongst spawners) was observed.

## DISCUSSION

Cantabrian populations, in common with other Atlantic salmon populations (Davidson et al., 1989), appear to have limited genetic variation at the protein level. Of the six loci screened, only two loci ( $M E P-2^{*}$ and $M D H-3,4^{*}$ ) were found to be polymorphic and the level of variation was small, typically with frequencies of the rarer allele $<0 \cdot 10$. Moreover, unlike other, larger Iberian salmon populations (Sánchez et al., 1991, 1996; Morán et al., 1994), salmon in Cantabrian rivers seem to be monomorphic for IDHP-3*, PGM-1*, LDH-4*, and GPI-1,2, though our sample sizes were small. Mitochondrial DNA variation, on the other hand, appears to be relatively high, and this probably reflects the ancient, pre-glacial origin of Atlantic salmon in the Iberian glacial refugium (Consuegra et al., 2002).

The populations studied appear to be stable over time at the two polymorphic protein loci examined, in some cases for up to 14 years. Temporal stability at protein loci is common in many Atlantic salmon populations (Crozier \& Moffett, 1989; Jordan et al., 1992; Moffett \& Crozier, 1996) and this has been associated with high levels of gene flow and large effective population sizes (Jordan et al., 1992). Temporal stability at protein loci could also indicate the existence of selection (Endler, 1986), however, as appears to be the case for the malic enzyme (MEP-2*) in Atlantic salmon (Verspoor \& Jordan, 1989; Jordan \& Youngson, 1991; Verspoor et al., 1991). If so, the temporal stability observed in Iberian populations may be more indicative of intense directional selection in extreme, marginal habitats, than of large population sizes and elevated levels of gene flow.

At a finer temporal scale, our results indicate that there is an important degree of seasonal structuring with respect to both phenotypic (body size, sea-age) and genetic (MEP-2*, mtDNA) traits. Body size and sea-age were found to decrease markedly as the season progressed, and both traits appeared to parallel the frequency of MEP-2* genotype. Thus, fish that were homozygous for the $M E P-2^{*}$ (100) allele remained longer at sea and tended to enter the rivers earlier than did fish with the * 125 allele, which in Cantabrian rivers were more likely to mature as grilse and entered the rivers later in the season. Fish that were homozygous for the *100 allele of $M E P-2^{*}$ were also significantly larger and heavier than * 125 homozygotes, while heterozygotes were intermediate in size. Such differences in body size, however, are largely due to the association of
Table VI. Predicted phenotypic and genetic changes that might occur in Cantabrian Atlantic salmon populations that result from the selective exploitation by anglers of early running fish (based on the observed differences between angler caught fish and spawners described in

| Trait | Predicted change | Observed change (1945-2000) | Reference |
| :---: | :---: | :---: | :---: |
| 1. Run timing | Shift towards delayed entry | Median timing of capture in the angling fishery delayed by 12-29 days | García de Leániz et al. (1992, 2001) |
| 2. Adult size | Reduction in body size | Average size of angler caught fish decreased by $2-6 \%$ in length and $10-15 \%$ in mass | García de Leániz et al. (1992, 2001) |
| 3. Age structure | Increased smolt age, lower sea-age, increased incidence of grilse | Widespread increase in the incidence of grilse in catch | García de Leániz et al. (1992, 2001) |
| 4. Life span | Reduced longevity | Reduced life span and almost complete loss of repeated spawners in all rivers | García de Leániz et al. $(1992,2001)$ |
| 5. $M E P-2^{*}$ | Reduced frequency of the common (*100) allele | Frequencies generally stable over time (1988-2000) | This publication (Table II) |
| 6. mtDNA ND1/16sRNA | Change in haplotypic frequencies | Significant temporal change in mtDNA haplotypic frequencies in the R. Asón | $\begin{aligned} & \text { Consuegra et al. } \\ & (2001,2002) \end{aligned}$ |

$M E P-2^{*}$ with age at maturity, which seems to determine the length of the marine growing season and, ultimately, the size attained by fish. Although the observed association between $M E P-2^{*}$ genotype and age at maturity has been reported previously in Atlantic salmon (Jordan et al., 1990; Morán et al., 1998), and is likely to be widespread, the implications of these differences for conservation have not been fully explored.

The observed differences in $M E P-2^{*}$ and mtDNA frequencies between angler caught fish and spawners strongly suggested that exploited fish are genetically, as well as phenotypically, distinct from spawners, hence the angling fishery exerts some artificial selection on the population. Moreover, if as suggested by some studies (Ballard \& Kreitman, 1995; Hey, 1997), mitochondrial genes are also affected by natural selection, then the differential removal by anglers of certain haplotypes may also disrupt the adaptive architecture of these populations (Consuegra et al., 2001, 2002). The selective exploitation of large fish may result in a lower fishery yield (Conover \& Munch, 2002) and also in a loss of genetic diversity and an impoverished gene pool as have been observed in other fish species (Law, 2000, 2001; Kenchington, 2003).

Atlantic salmon populations subjected to the kind of seasonal exploitation exerted by anglers in Iberian rivers may be expected to change in several ways (Table VI). First, a shift towards delayed entry is likely to occur since timing of entry into the rivers is inherited (Hansen \& Jonsson, 1991; Stewart et al., 2002) and early running fish are exploited by the fishery while late running fish are not. Likewise, reductions in body size, sea-age and longevity are expected to occur, as these traits are also under genetic control in Atlantic salmon (Taylor, 1991; Rye \& Refstie, 1995). Finally, genetic changes may also take place if spawners and angler caught fish differ with respect to $M E P-2^{*}$ and mtDNA. Many of these predicted changes have been detected in Cantabrian populations (Table VI), indicating that angling for early running fish may induce detrimental genetic and phenotypic changes in Atlantic salmon populations.

The results suggest that early and late runs of Atlantic salmon constitute demographically and genetically distinct components of Iberian Atlantic salmon populations and that the selective exploitation of early running adults by angling may alter the adaptive architecture of these populations. It is recommended that the seasonal dimension is incorporated into the definition of operational conservation units for Atlantic salmon (Dodson et al., 1998) and that rod and line fisheries are managed to reduce the risk of selective harvesting. Failure to do so may mean that well intentioned fishery regulations, such as fishing closures, may inadvertently compromise the same salmon populations they were designed to protect.

[^1]
## References

Adams, N. S., Spearman, W. J., Burger, C. V., Currens, K. P., Schreck, C. B. \& Hiram, W. L. (1994). Variation in mitochondrial DNA and allozymes discriminates early
and late forms of chinook salmon (Oncorhynchus tshawytscha) in the Kenai and Kasilof Rivers, Alaska. Canadian Journal of Fisheries and Aquatic Sciences 51, 172-181.
Ballard, J. W. O. \& Kreitman, M. (1995). Is mitochondrial DNA a strictly neutral marker? Trends in Ecology and Evolution 10, 485-488.
Caswell, H., Naiman, R. J. \& Morin, R. (1984). Evaluating the consequences of reproduction in complex salmonid life cycles. Aquaculture 43, 123-134.
Clayton, J. W. \& Tretiak, D. N. (1972). Amine-citrate buffer for pH control in starch gel electrophoresis. Journal of the Fisheries Research Board of Canada 29, 1169-1172.
Conover, D. O. \& Munch, S. B. (2002). Sustaining fisheries yields over evolutionary time scales. Science 297, 94-96.
Consuegra, S., García de Leániz, C., Serdio, A., Knox, D. \& Verspoor, E. (2001). Conservation genetics of endangered Atlantic salmon populations: the River Asón, a case study. In El Salmón, Joya de Nuestros Ríos (García de Leániz, C., Serdio, A. \& Consuegra, S., eds), pp. 213-226. Santander: Consejería de Ganadería, Agricultura y Pesca (in Spanish with English summary).
Consuegra, S., García de Leániz, C., Serdio, A., González Morales, M., Straus, L. G., Knox, D. \& Verspoor, E. (2002). Mitochondrial DNA variation in Pleistocene and modern Atlantic salmon from the Iberian glacial refugium. Molecular Ecology 11, 2037-2048.
Cronin, M. A., Spearman, W. J., Wilmot, R. L., Patton, J. C., \& Bickman, J. W. (1993). Mitochondrial DNA variation in chinook (Oncorhynchus tshawytscha) and chum salmon ( O. keta) detected by restriction enzyme analysis of polymerase chain reaction (PCR) products. Canadian Journal of Fisheries and Aquatic Sciences 50, 708-715.
Crozier, W. W. \& Moffett, I. J. J. (1989). Amount and distribution of biochemicalgenetic variation among wild and a hatchery stock of Atlantic salmon, Salmo salar L., from north-east Ireland. Journal of Fish Biology 35, 665-677.

Davidson, W. S., Birt, T. P. \& Green, J. M. (1989). A review of genetic variation in Atlantic salmon, Salmo salar L., and its importance for stock identification, enhancement programmes and aquaculture. Journal of Fish Biology 34, 547-560.
Dodson, J. J., Gibson, R. J., Cunjak, R. A., Friedland, K. D., García de Leániz, C., Gross, M. R., Newbury, R., Nielsen, J. L., Power, M. E. \& Roy, S. (1998). Elements in the development of conservation plans for Atlantic salmon (Salmo salar). Canadian Journal of Fisheries and Aquatic Sciences 55, 312-323.
Endler, J. A. (1986). Natural Selection in the Wild. Princeton, NJ: Princeton University Press.
Ferguson, A., Taggart, J. B., Prodöhl, P. A., McMeel, O., Thompson, C., Stone, C., McGinnity, P. \& Hynes, R. A. (1995). The application of molecular markers to the study and conservation of fish populations, with special reference to Salmo. Journal of Fish Biology 47, 103-126.
García de Leániz, C. \& Martínez, J. J. (1988). The Atlantic Salmon in the Rivers of Spain with particular reference to Cantabria. In The Atlantic Salmon: Planning for the Future (Mills, D. \& Piggins, D., eds), pp. 179-209. London: Croom-Helm \& Timber Press.
García de Leániz, C., Verspoor, E. \& Hawkins, A. D. (1989). Genetic determination of the contribution of stocked and wild Atlantic salmon, Salmo salar L., to the angling fisheries in two Spanish rivers. Journal of Fish Biology 35, 261-270.
García de Leániz, C., Caballero, P., Valero, E., Martínez, J. J. \& Hawkins, A. D. (1992). Historical changes in Spanish Atlantic salmon (Salmo salar L.) rod and line fisheries: why are large multi-seawinter fish becoming scarcer? Journal of Fish Biology (Supplement B) 41, 179 (Abstract).
García de Leániz, C., Serdio, A. \& Consuegra, S. (2001). Present status of Atlantic salmon in Cantabria. In El Salmón, Joya de Nuestros Ríos (García de Leániz, C., Serdio, A. \& Consuegra, S., eds), pp. 55-82. Santander: Consejería de Ganadería, Agricultura y Pesca (in Spanish with English summary).

Gardner, M. L. G. (1976). A review of factors which may influence the sea-age of maturation of Atlantic salmon Salmo salar L. Journal of Fish Biology 9, 289-327.
Gee, A. S. \& Milner, N. J. (1980). Analysis of 70-year catch statistics for Atlantic salmon (Salmo salar) in the River Wye and implications for management of stocks. Journal of Applied Ecology 17, 41-57.
Hansen, L. P. \& Jonsson, B. (1991). Evidence of a genetic component in the seasonal return patterns of Atlantic salmon, Salmo salar L. Journal of Fish Biology 38, 251-258.
Hey, J. (1997). Mitochondrial and nuclear genes present conflicting portraits of human origins. Molecular Biology and Evolution 14, 166-172.
Hutchings, J. A. (2001). Influence of population decline, fishing, and spawner variability on the recovery of marine fishes. Journal of Fish Biology 59, 306-322.
Jordan, W. C. \& Youngson, A. F. (1991). Genetic protein variation and natural selection in Atlantic salmon (Salmo salar L.) parr. Journal of Fish Biology 39, 185-192.
Jordan, W. C., Youngson, A. F. \& Webb, J. H. (1990). Genetic variation at the malic enzyme-2 locus and age at maturity in sea-run Atlantic salmon (Salmo salar). Canadian Journal of Fisheries and Aquatic Sciences 47, 1672-1677.
Jordan, W. C., Youngson, A. F., Hay, D. W. \& Ferguson, A. (1992). Genetic protein variation in natural populations of Atlantic salmon (Salmo salar) in Scotland: temporal and spatial variation. Canadian Journal of Fisheries and Aquatic Sciences 49, 1863-1972.
Kenchington, E. L. (2003). The effects of fishing on species and genetic diversity. In Responsible Fisheries in the Marine Ecosystem (Sinclair, M. \& Valdimarsson, G., eds), pp. 235-253. CABI Publishing.
Law, R. (2000). Fishing, selection, and phenotypic evolution. ICES Journal of Marine Science 57, 659-668.
Law, R. (2001). Phenotypic and genetic changes due to selective exploitation. In Conservation of Exploited Species (Reynolds, J. D., Mace, G. M., Redford, K. H. \& Robinson, J. G., eds), pp. 323-342. Cambridge: Cambridge University Press.
McElroy, D., Moran, P., Bermingham, E. \& Kornfield, I. (1991). REAP: the Restriction Enzyme Analysis Package. (4.0). Orono, Maine, Department of Zoology, University of Maine.
Moffett, I. J. J. \& Crozier, W. W. (1996). A study of temporal genetic variation in a natural population of Atlantic salmon in the River Bush, Northern Ireland. Journal of Fish Biology 48, 302-306.
Morán, P., Pendás, A. M., García-Vázquez, E. \& Izquierdo, J. I. (1994). Genetic variation among Atlantic salmon in six Spanish Rivers. Journal of Fish Biology 45, 831-837.
Morán, P., Pérez, J. \& García-Vázquez, E. (1998). The malic enzyme MEP-2 locus in Spanish populations of Atlantic salmon: sea age and foreign stocking. Aquatic Sciences 60, 359-366.
Palumbi, S. R. (1996). Nuclear Acids II: The polimerase chain reaction. In Molecular Systematics (Hillis, D. M., Moritz, C. \& Mable, B. K., eds), pp. 205-247. Sunderland: Sinauer Associates.
Policansky, D. (1993). Fishing as a cause of evolution in fisheries. In The Exploitation of Evolving Resources (Stokes, T. K., McGlade, J. M. \& Law, R., eds), pp. 2-18. Lectures Notes in Biomathematics 99, Heidelberg: Springer-Verlag.
Raymond, M. \& Rousset, F. (1995). GENEPOP (vers. 1.2): Population genetics software for exact tests and ecumenicism. Journal of Heredity 86, 248-249.
Rye, M. \& Refstie, T. (1995). Phenotypic and genetic parameters of body size traits in Atlantic salmon (Salmo salar L.). Aquaculture Research 26, 875-885.
Sánchez, J. A., Blanco, G., Vazquez, E., García, E. \& Rubio, J. (1991). Allozyme variation in natural populations of Atlantic salmon in Asturias (northern Spain). Aquaculture 93, 291-298.
Sánchez, J. A., Clabby, C., Ramos, D., Blanco, G., Flavin, F., Vázquez, E. \& Powell, R. (1996). Protein and microsatellite single locus variability in Salmo salar L. (Atlantic salmon). Heredity 77, 423-432.

Shaklee, J. B., Allendorf, F. W., Morizot, D. C. \& Whitt, T. S. (1989). Genetic nomenclature for protein-coding loci in fish: proposed guidelines. Transactions of the American Fisheries Society 118, 218-227.
Shearer, W. M. (1992). The Atlantic Salmon. Natural History, Exploitation and Future Management. London: Fishing News Books.
Smith, P. M., Francis, R. I. C. C. \& McVeigh, M. (1991). Loss of genetic diversity due to fishing pressure. Fisheries Research 10, 309-316.
Stewart, D. C., Smith, G. W. \& Youngson, A. F. (2002). Tributary-specific variation in timing of return of adult Atlantic salmon (Salmo salar) to fresh water has a genetic component. Canadian Journal of Fisheries and Aquatic Sciences 59, 276-281.
Taggart, J. B., Hynes, R. A., Prodöhl, P. A. \& Ferguson, A. (1992). A simplified protocol for routine total DNA isolation from salmonid fishes. Journal of Fish Biology 40, 963-965.
Taylor, E. B. (1991). A review of local adaptations in Salmonidae, with particular reference to Pacific and Atlantic salmon. Aquaculture 98, 185-207.
Thorpe, J. E. (1993). Impacts of fishing on genetic structure of salmonid populations. In Genetic Conservation of Salmonid Fishes (Cloud, J. G. \& Thorgaard, G. H., eds), pp. 67-80. New York: Plenum Press.
Verspoor, E. (1988). Reduced genetic variability in first-generation hatchery populations of Atlantic salmon (Salmo salar). Canadian Journal of Fisheries and Aquatic Sciences 45, 1686-1690.
Verspoor, E. (1994). The evolution of genetic divergence at protein coding loci among anadromous and non-anadromous populations of Atlantic salmon (Salmo salar). In Genetics and Evolution of Aquatic Organisms (Beaumont, A. R., ed.), pp. 52-67. London: Chapman \& Hall.
Verspoor, E. \& Cole, L. J. (1989). Genetically distinct sympatric populations of resident and anadromous Atlantic salmon, Salmo salar. Canadian Journal of Zoology 67, 1453-1461.
Verspoor, E. \& Jordan, W. C. (1989). Genetic variation at the Me-2 locus in the Atlantic salmon within and between rivers: evidence for its selective maintenance. Journal of Fish Biology 35, 205-213.
Verspoor, E., Fraser, N. H. C. \& Youngson, A. F. (1991). Protein polymorphism in Atlantic salmon within a Scottish river: evidence for selection and estimates of gene flow between tributaries. Aquaculture 98, 217-230.
Verspoor, E., McCarthy, E. M., Knox, D., Bourke, E. \& Cross, T. F. (1999). The phylogeography of European Atlantic salmon (Salmo salar L.) based on RFLP analysis of the ND1/16sRNA region of the mtDNA. Biological Journal of the Linnean Society 68, 129-146.


[^0]:    $\$$ Author to whom correspondence should be addressed. Tel.: +44 (0) 1792 295383; fax: +44 (0) 1792 295447; email: c.garciadeleaniz@swansea.ac.uk

[^1]:    The authors are grateful to the Cantabrian wildlife bailiffs for collecting most of the samples, to the personnel and students at Centro Ictiológico de Arredondo for help with the study, and to two anonymous referees for helpful comments. This work was funded by the Cantabrian Regional Government (Consejería de Ganadería, Agricultura y Pesca, Dirección General de Montes y Conservación de la Naturaleza) and Scottish Executive Fisheries Research Service.

